Biology, diagnosis and treatment of canine appendicular osteosarcoma: Similarities and differences with human osteosarcoma

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ABSTRACT

Osteosarcoma (OSA) is the most common primary bone tumour in dogs. The appendicular locations are most frequently involved and large to giant breed dogs are commonly affected, with a median age of 7-8 years. OSA is a locally invasive neoplasm with a high rate of metastasis, mostly to the lungs. Due to similarities in biology and treatment of OSA in dogs and humans, canine OSA represents a valid and important tumour model. Differences between canine and human OSAs include the age of occurrence (OSA is most commonly an adolescent disease in humans), localisation (the stifle is the most common site of localisation in humans) and limited use of neoadjuvant chemotherapy in canine OSA.

Introduction

Spontaneous tumours in dogs may serve as models for human cancer biology and translational cancer therapeutics, having greater similarity than many current experimental tumour models. Canine osteosarcoma (OSA) is a suitable model for OSA in humans due to the relatively high incidence of the tumour in dogs, similarities in biological behaviour, common molecular features, large body size of breeds more frequently affected and sharing of the same environment. The lack of specific chemotherapeutic drugs in veterinary medicine and the sometimes prohibitive costs of treatment for the management of cancer in dogs allow early access to novel therapeutics. Dogs have a naturally shorter life span, with more rapid progression and early metastatic failure of cancer, permitting more rapid completion of clinical trials in this species compared to human patients. This paper gives an overview of the biology and treatment options of OSA in dogs and humans.

Clinical presentation of appendicular osteosarcoma

Canine OSA accounts for 85-98% of all canine bone tumours (Liptak et al., 2004b; Dernell et al., 2007). Appendicular locations are more common (75%), but OSA can also affect the axial skeleton (24%) and, occasionally, soft tissues (1%). Appendicular OSA is a locally aggressive malignant neoplasm which destroys bone locally, extends into surrounding soft tissues and has a high rate of metastasis. Metastasis occurs mainly to the lungs via the haematogenous route, as well as to other bones, visceral organs, the brain, subcutaneous tissue and skin (Gorman et al., 2006; Dernell et al., 2007). Lymph nodes are involved less commonly, with reported frequencies of 4.4-9.0% (Brodey and Abt, 1976; Spodnick et al., 1992; Hillers et al., 2005). The use of chemotherapy as part of standard curative-intent treatment is associated with an increase in the rate of bone and soft tissue metastases (Dernell et al., 2007).

In dogs, appendicular OSAs most often affect the metaphyses of long bones. The fore limbs are affected twice as often as the hind limbs, with the distal radius and proximal humerus being the most frequent sites, followed by the distal femur and proximal and distal tibia (Straw et al., 1990; Dernell et al., 2007). Large and giant breeds are more commonly affected; only 5% of OSAs occur in dogs <15 kg (Dernell et al., 2007). Greyhounds, Rotweilers, Great Danes, Saint Bernards, Doberman Pinschers, Irish Setters, Golden Retrievers and German Shepherds have an increased risk of developing OSA, even though the predisposition seems to be related to size rather than breed (Ru et al., 1998; Rosenberger et al., 2007). A familial pattern of
occurrence has been observed in the Saint Bernard, Rottweiler and Scottish Deerhound (Misdorp, 1980; Phillips et al., 2007). Males are more often affected than females (ratio 1.5:1), but this finding is not consistent among publications (Brodey and Abt, 1976; Mauldin et al., 1988; Shapiro et al., 1988; Kraegel et al., 1991; Spodnick et al., 1992). Affected females mainly belong to the Great Dane, Saint Bernard and Rottweiler breeds. The age at presentation has a bimodal distribution; a first peak is reported at 18-24 months, but most dogs are 7-9 years old (Dernell et al., 2007). Neutered dogs have twice the risk of developing OSA com-pared with sexually intact dogs (Ru et al., 1998). A study on 683 Rottweilers undergoing gonadectomy before 1 year of age found a strong inverse association between lifetime exposure to gonadal hormones and incidence of OSA, suggesting that sex hormones may play a role in tumour development (Cooley et al., 2002).

Human OSA is the most common primary solid bone tumour in childhood and adolescence (Carriere and Bielack, 2006; Ta et al., 2009). The incidence is higher in the second decade of life, during periods of rapid bone turnover, with a median age of 16 years (Ta et al., 2009). After 10 years of age, males are more frequently affected than females. A second peak in incidence occurs in older patients, usually associated with underlying bone pathology, such as Paget’s disease, medullary infarcts or prior irradiation (Ta et al., 2009). The metaphysis of long bones is the primary site in more than 80% of cases. OSAs most commonly present at sites of rapid bone turnover, such as the distal femur, proximal tibia and proximal humerus (Messerschmitt et al., 2009).

Metastases are clinically detectable in approximately 20% of human patients on initial presentation (O’Day and Gorlick, 2009) and metastatic spread is usually by the haematogenous route. The lungs are the most common metastatic site (80-85%), followed by bone (10%), which is usually involved only after pulmonary metastasis (Federman et al., 2009). Less frequent sites of metastasis include lymph nodes (<10%), liver, adrenal glands, central nervous system, muscle and skin (Ta et al., 2009). Patients without clinically detectable metastases are presumed to have micrometastatic disease (Messerschmitt et al., 2009).

**Aetiology and risk factors for osteosarcoma**

The aetiology of most OSAs remains unknown both in humans and dogs. Some factors have been identified as possibly being involved in development of canine OSA.

**Dogs**

**Ionising radiation**

In both therapeutic and experimental settings, exposure to ionising radiation can induce OSA. Beagles administered aerosols containing plutonium dioxide developed OSAs in the lungs, skeleton and liver, beginning about 3 years after exposure (Mugenburg et al., 1996). Skeletal malignancies, most of which were OSAs, were documented among 234 young adult beagles given single intravenous injections of monomeric 239Plutonium citrate (Lloyd et al., 1993). In another experimental study, 36/117 young adult beagles injected with 239Americium developed OSA (Lloyd et al., 1994).

Several reports of OSA as a late complication of radiation therapy in dogs have been described. A vertebral OSA occurred in a dog 5 years after 60cobalt teletherapy for a spinal cord tumour (Dickinson et al., 2001). Secondary OSAs developed within the field of megavoltage irradiation 1.7-5 years after treatment in 3/87 (3.4%) of spontaneous tumour-bearing dogs irradiated for soft tissue sarcomas (Gillette et al., 1990). OSA has also been reported after orthovoltage irradiation of oral acanthomatous epulis (Thrall, 1984; White et al., 1986). In an experimental study, 21% of dogs undergoing intra-operative radiation therapy (>25 Gy) to the vertebral column, followed in some cases by external beam radiation, developed OSA 4-5 years post-treatment (Powers et al., 1989).

**Minor chronic trauma**

Long-standing metallic implants (e.g. Jonas intramedullary splints and older generation tibial plateau levelling osteotomy plates) after orthopaedic procedures have been associated with the development of OSA (Sinibaldi et al., 1976, 1982; Murphy et al., 1997; Boudrieau et al., 2005; Harasen and Simko, 2008). Hypotheses to explain this phenomenon include a direct effect of metal implants, infection, instability of the implant and corrosion (Stevenson, 1991). However, given the large number of orthopaedic surgical implants routinely applied and the fact that no conclusions have been drawn on the role of metallic implants in sarcoma development, the occurrence of malignant lesions at the same site may be no more than a coincidence (Murphy et al., 1997). Underlying diseases, such as spontaneous or post-orthopaedic surgery bone infarcts and osteochondritis dissecans, have also been reported as possible causative factors in dogs (Riser et al., 1972; Dubielzig et al., 1981; Marcellin-Little et al., 1999; Holmberg et al., 2004). A study by Gellasch et al. (2002) on two groups of dogs of different sizes (<15 kg; >25 kg) failed to demonstrate increased microdamage in the distal metaphyseal radius in the large size group, suggesting that microdamage is unlikely to be an important risk factor for OSA.

**Genetic alterations**

In one study, 27 p53 tumour suppressor gene mutations (20 point mutations and 7 deletions) were observed in 24/59 (40.7%) canine OSAs (Kirpensteijn et al., 2008). Cases of OSA with mutated p53 had a decreased survival time compared to dogs without p53 alterations (Kirpensteijn et al., 2008). In two other studies, p53 was over-expressed in the majority of canine OSAs and alterations in its expression correlated with highly aggressive tumour behaviour and higher tumour grade (Sagartz et al., 1996; Loukopoulos et al., 2003). Mutations in p53 have also been observed in other studies of canine OSA by van Leeuwen et al. (1997) and Mendoza et al. (1998).

Over-expression of erb-B2, which encodes human epidermal growth factor receptor 2 (HER-2), was observed in 86% and
40% of canine OSA cell lines and tissue samples, respectively (Flint et al., 2004). Deletions, mutations and down-regulation of the PTEN tumour suppressor gene have also been detected in OSA cell lines and tumour samples (Levine et al., 2002). Hepatocyte growth factor (HGF) and its receptor c-Met were expressed in most OSA samples (Ferracini et al., 2000; De Maria et al., 2009; Fieten et al., 2009). A role for insulin-like growth factor-1 (IGF-1) and its receptor (IGF-1R) in cell growth and invasion in OSA canine cell lines has been demonstrated (MacEwen et al., 2004). Matrix metalloproteinases 2 and 9, which may contribute to local disease progression and metastatic spread, are expressed in OSA cell lines and tissues (Lana et al., 2000; Loukopoulos et al., 2004). Similarly, ezrin, a membrane cytoskeleton linker also potentially involved in metastasis, was detected in 83% of primary canine OSAs and its presence was associated with a shorter median disease-free interval compared to OSAs with low ezrin expression (Khanna et al., 2004). Constitutive activation of signal transducer and activator of transcription 3 (STAT3) was present in a subset of canine OSA tumours and cell lines, but not in normal canine osteoblasts (Fossey et al., 2009).

Humans

Factors associated with the development of OSA in humans include the faster growth rate of bone at puberty, exposure to chemicals such as beryllium, cytogenetic abnormalities, hereditary retinoblastoma (mutations in the RB gene), Li-Fraumeni syndrome (mutations in the p53 gene), Bloom syndrome, Rothmund-Thomson syndrome and Werner's syndrome (Mueller et al., 2007; Ta et al., 2009). Radiation-induced OSAs are rare and typically occur in adults because of the long interval (5-20 years) between radiation exposure and neoplastic transformation (Mala-Weer et al., 2008; Ta et al., 2009).

In human OSAs, tumorigenesis has been associated with alterations in tumour suppressor proteins (p53, Rb, PTEN), alterations in oncogene expression (erbB-2, MET) and dysregulated cell signalling and kinase pathways, such as vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), mTOR, c-Kit, metalloproteinases and ezrin (Mueller et al., 2007; O’Day and Gorlick, 2009; Ta et al., 2009).

History and physical examination

Dogs

Dogs with appendicular OSA are referred for the onset of progressive lameness and leg swelling (Dernell et al., 2007). Lameness is usually intermittent and initially mild, but progresses to become persistent and severe. The mass at the primary site is usually firm and often painful on palpation. Acute non-weight bearing lameness is typically associated with a pathological fracture.

Humans

Human patients with OSA present with pain of several months’ duration (2-4 months before diagnosis), usually related to strenuous exercise or trauma, and the pain interferes with sleep. On clinical examination, a visible swelling with a hard painful mass, decreased joint mobility or localised warmth or erythema may be present. Approximately 5-10% of patients with OSA present with a pathological fracture (Messerschmitt et al., 2009; Ta et al., 2009).

Diagnosis and staging

Dogs

A diagnosis of primary malignant bone tumour is often suggested by clinical presentation and radiographic findings. Initial diagnosis can be attempted by fine needle aspiration and cytology (Loukopoulos et al., 2005; Reinhardt et al., 2005; Britt et al., 2007). Alkaline phosphatase (ALP) staining of cytology samples is useful for differentiating between OSA and other primary bone tumours (Britt et al., 2007). Bone biopsy can be performed via closed (Jamshidi needle or Michelle’s trephine) or open techniques (Powers et al., 1988). The diagnostic quality of cytological or histological bone samples can be improved by image-guided techniques (Vignoli et al., 2004; Britt et al., 2007).

Diagnostic imaging plays an important role in diagnosis and staging of dogs with OSA. Cranio-caudal and latero-medial radiographic views of the primary lesion, including the joint above and below the affected bone, are required. The radiographic appearance of OSA in long bones includes cortical bone lysis and/or a proliferative sunburst pattern, periosteal proliferation, sub-periosteal new bone formation and soft tissue swelling, with calcification extending into surrounding soft tissue.

Several studies have been performed to evaluate the accuracy of radiography, bone scintigraphy, computed tomography (CT) and magnetic resonance imaging (MRI) in assessing the local extent of appendicular OSA (LaRue et al., 1986; Berg et al., 1990; Hahn et al., 1990). In one study, MRI was recognised as the best modality for preoperative assessment of intramedullary extent of appendicular OSA when limb-sparing was an option (Wallach et al., 2002). Another study comparing radiographs, MRI and CT in 10 post-amputation OSA cases failed to identify a superior modality in predicting extent of tumour infiltration (Davis et al., 2002). Nuclear scintigraphy overestimated the local extent of OSA more than radiography, providing a larger
margin of safety when determining the site of the proximal osteotomy, but also decreasing candidates for limb-sparing surgery (Leibman et al., 2001).

Thoracic radiographs are performed to evaluate metastatic spread to the lungs. Only a few patients (<5-10%) are positive for radiographic lung metastasis at presentation, but OSA is considered to be a tumour with a high metastatic potential, since approximately 90% of dogs with OSA treated by amputation only die of metastatic disease, usually to the lung, within 1 year of diagnosis (Dernell et al., 2007). CT of the thorax is superior to radiography in detecting smaller lung lesions (Nemanic et al., 2006). Using nuclear scintigraphy, the incidence of occult bone metastasis at the time of diagnosis of the primary tumour is 7.9%; lesions appear as non-specific areas of increased uptake of radiopharmaceutical that should be verified by radiographs or biopsy (Jankowski et al., 2003).

**Humans**

In humans, at least two orthogonal radiographic views are required when a bone lesion is suspected. The classical radiographic appearance is of ill-defined borders, an osteoblastic and/or osteolytic lesion and an associated soft tissue mass. MRI represents the primary mode of evaluation of OSA in humans and can clearly demonstrate the extent of tumour invasion of the surrounding soft tissue, neurovascular involvement, extent of bone marrow replacement and presence of discontinuous metastases (Federman et al., 2009). MRI is also useful to assess the possibility of limb salvage. CT-guided core biopsy is frequently used for tissue biopsy for histopathological diagnosis (Federman et al., 2009). A CT scan of the chest and a nuclear scintigraphy bone scan are recommended to rule out metastasis to the lungs and bone. Interest in the use of positron-emission tomography (PET) for staging OSAs and monitoring treatment is increasing (Federman et al., 2009).

**Treatment**

**Dogs**

Dogs with appendicular OSA can be managed with either palliative- or curative-intent therapy. Curative-intent treatment is aimed at local tumour control and prevention or delay of metastatic disease.

**Limb amputation**

Limb amputation remains the current standard of care for local management (Brodey and Abt, 1976; Spodnick et al., 1992). It avoids the risk of pathological fracture, eliminates pain and is a well tolerated procedure, with minimal complications; even large breed dogs show good functional results and the majority of owners are satisfied with the pet's quality of life (Carberry and Harvey, 1986; Kirpensteijn et al., 1999). Contraindications to amputation may be severe obesity or concurrent debilitating orthopaedic or neurological diseases; however, each case needs to be evaluated on an individual basis.

**Limb-sparing surgery**

Limb-sparing in dogs can be achieved with surgical and/or radiation techniques. Good results have been reported with reconstructive limb-sparing procedures for OSA of the distal radius, whereas limb-sparing surgery at other sites is associated with a higher complication rate and poor limb function (Kuntz et al., 1998; Morello et al., 2001). Conversely, a lower complication rate and good restoration of limb function have been reported after ablative limb-sparing techniques for OSA in the ulna, scapula, metacarpus, metatarsus and ischium.

Several surgical techniques have been used to preserve the limb and they represent a valid alternative to amputation. After surgical resection of the OSA, the defect may be filled with a frozen cortical allograft (LaRue et al., 1989; Morello et al., 2001; Liptak et al., 2004a), an endoprosthesis (Liptak et al., 2006) (Fig. 1) or with the resected neoplastic bone after it has been pasteurised (Buracco et al., 2002; Morello et al., 2003) (Fig. 2), autoclaved (Massin et al., 1995; Yamamoto et al., 2002) or irradiated (Yamamoto et al., 2002; Liptak et al., 2004c; Boston et al., 2007). Bone or metallic implants are fixed in position with a bone plate and screws, and arthrodesis of the adjacent joint is performed. A combination of allograft and prosthesis has been used to preserve the limb for OSA of the proximal femur (Liptak et al., 2005). The more common complications associated with cortical allograft, pasteurised autograft and endoprosthesis include local tumour recurrence (15-28%), infection (31-60%) and implant failure or loosening (11-40%) (LaRue et al., 1989; Straw and Withrow, 1996; Morello et al., 2001, 2003; Liptak et al., 2006).

After excision of OSAs from the distal radius or tibia, large surgical defects have been replaced by vascularised, viable, regenerated bone by single (Fig. 3) or double transport osteogenesis (Tommasini-Degna et al., 2000; Rovesti et al., 2002; Ehrhart, 2005). Transverse ulnar bone transport osteogenesis has also been investigated experimentally (Jehn et al., 2007). These procedures can achieve good limb function and absence of infection, but problems include local tumour recurrence, owner compliance in distracting the apparatus several times per day and apparatus failure. Limb-sparing surgery has also been performed by rolling the distal ulna into the distal radial defect after tumour excision, thus using the ulna as a vascularised transposition autograft (Seguin et al., 2003). However, the procedure is more likely to have biomechanical complications compared to the standard cortical allograft technique (Pooya et al., 2004).
Surgery alone is considered to be palliative. The reported mean survival time after surgery alone is 103-175 days (Brodey and Abt, 1976; Mauldin et al., 1988; Shapiro et al., 1988; Straw et al., 1991; Thompson and Fugent, 1991; Berg et al., 1992; Spodnick et al., 1992). The 1- and 2-year survival is 11-20% and 2-4%, respectively (Straw et al., 1991; Thompson and Fugent, 1991; Berg et al., 1992; Spodnick et al., 1992). There are no statistical differences in survival between amputation and limb-sparing surgery if
adequate systemic chemotherapy is given (Straw and Withrow, 1996). An improvement in survival with limb-sparing is only evident if the surgical field becomes infected; the median survival time for dogs with infected surgical sites after limb-sparing is 685 days compared to 289 days in the absence of infection (Lascelles et al., 2005; Liptak et al., 2006). Similar findings have also been reported in humans with limb-sparing surgery (Jeys et al., 2007).

**Chemotherapy**

Adjuvant chemotherapy can improve survival of dogs with OSA when associated with surgery and/or radiotherapy. Chemotherapy protocols include doxorubicin, cisplatin, carboplatin and lobaplatin used alone or in combination (Table 1). A local cisplatin delivery system has been described (Straw et al., 1994; Withrow et al., 2004; Mehl et al., 2005). Administration of chemotherapy in addition to surgery and/or radiotherapy increases the median survival time from 103-175 days to 262-450 days. The 1- and 2-year survival rates with chemotherapy range from 31-48% to 10-26%, respectively. Survival times for dogs treated with single agent platinum compounds are similar to those reported with combined protocols (Dernell et al., 2007). The most efficacious chemotherapeutic agent and the ideal timing to start adjuvant chemotherapy have not been identified. However, there is no substantial advantage in early post-operative chemotherapy (Berg et al., 1997; Dernell et al., 2007), so it is better to wait an adequate time to allow the patient to recover from surgery and early healing of the surgical wound (Berg et al., 1997). Chemotherapy is usually less effective in the presence of macroscopic metastatic disease (Ogilvie et al., 1993). The efficacy of aerosol-delivered gemcitabine has been investigated by Rodriguez et al. (2010) in dogs with pulmonary metastatic OSA. Pulmonary metastatectomy resulted in significantly prolonged survival in selected patients (O’Brien et al., 1993).

**Fig. 3.** Single bone transport osteogenesis limb-sparing surgery in the distal radius of a dog with osteosarcoma. Post-operative image of the circular fixator (a). Post-operative lateral (b) and cranio-caudal (c) radiographs of distal radius.

**Table 1.** Chemotherapeutic agents used for dogs with appendicular osteosarcoma.
**Immunotherapy**

Immunotherapy has been combined with chemotherapy to enhance anti-tumour effects without increasing toxicity. Anti-tumour activity was observed when the immunomodulatory agent L-muramyl tripeptide-phosphatidylethanolamine (L-MTP-PE) was administered to dogs with OSA after limb amputation or limb-sparing surgery and completion of cisplatin treatment (MacEwen et al., 1994; Kurzman et al., 1995). L-MTP-PE administered with doxorubicin enhanced canine monocyte activation induced by doxorubicin or L-MTP-PE alone (Shi et al., 1993) and induced cytotoxic activity of pulmonary alveolar macrophages against OSA cells when compared to dogs treated with doxorubicin or L-MTP-PE alone (Kurzman et al., 1999).

**Radiotherapy**

Radiation therapy has a role in curative-intent local treatment of canine appendicular OSA. Walter et al. (2005) proposed a full-course fractionated external beam protocol in conjunction with cisplatin, but there was no substantial improvement over a palliative protocol. A single fraction of 70 Gy given intra-operatively after exteriorisation of the tumoral bone segment has been used in combination with chemotherapy (Liptak et al., 2004c; Boston et al., 2007). Post-operative complications may be high (69%) and include deep infection, fracture of irradiated bone, implant failure and local recurrence. This procedure should only be performed in dogs bearing appendicular OSAs at sites in which limb-sparing techniques are not an option.

A stereotactic radiosurgery protocol has also been used, in which dogs are irradiated with a single large targeted dose (30-50 Gy), with or without chemotherapy (Farese et al., 2004). This allows normal tissue to be spared and avoids the need for surgery in some cases, although pathological fractures may occur.

**Palliative-intent treatment**

The aim of palliative-intent treatment is to alleviate pain. Radiation therapy is a valid method of palliation for appendicular OSA, inducing relief of pain, reduced lameness and improving quality of life, whereas radiation-induced acute side effects are rare. Radiation therapy protocols include a two-fraction protocol (Ramirez et al., 1999), three-fraction protocols (Bateman et al., 1994; Ramirez et al., 1999; Mueller et al., 2005), four-fraction protocol (Green et al., 2002) and expedited protocol (Knapp-Hoch et al., 2009). The effectiveness and duration of analgesia among these protocols range from 50-93% and 53-180 days, respectively. Most dogs died or were euthanased because of local disease progression, metastatic disease or pathological fractures.

It is not clear if dogs with OSA benefit from more durable pain relief when chemotherapy is combined with radiotherapy (Ramirez et al., 1999; Mueller et al., 2005). Boston et al. (2006) re-reported that longer survival (median survival time 130 days) can be achieved in dogs with metastatic (stage III) appendicular OSA using palliative radiation compared with surgery alone. Palliative-intent treatment for canine OSA has also been attempted by intravenous administration of $^{153}$Samarium (Barnard et al., 2007).

Bisphosphonates have been proposed for palliative-intent treatment in dogs with OSA. Clinical applications include therapy for tumour-related hypercalcaemia, inhibition of bone metastasis and pain relief. Zoledronic acid (Spugnini et al., 2009), pamidronate (Fan et al., 2005, 2007) and alendronate (Tomlin et al., 2000) provided pain palliation in some treated dogs.

<table>
<thead>
<tr>
<th>Drug</th>
<th>Median survival time (days)</th>
<th>1-year survival (%)</th>
<th>2-year survival (%)</th>
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<tr>
<td>Cisplatin</td>
<td></td>
<td></td>
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<tr>
<td>Thompson and Fugent (1991)</td>
<td>250</td>
<td>33</td>
<td>NR*</td>
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<tr>
<td>Shapiro et al. (1988)</td>
<td>301</td>
<td>NR</td>
<td>NR</td>
</tr>
<tr>
<td>Straw et al. (1991)</td>
<td>262</td>
<td>38</td>
<td>18</td>
</tr>
<tr>
<td>Kraegel et al. (1991)</td>
<td>413</td>
<td>62</td>
<td>NR</td>
</tr>
<tr>
<td>Berg et al. (1992)</td>
<td>325</td>
<td>45.5</td>
<td>20.9</td>
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<tr>
<td>Cisplatin + doxorubicin</td>
<td></td>
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<tr>
<td>Chan et al. (2005)</td>
<td>300</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>Mauldin et al. (1988)</td>
<td>300</td>
<td>36.8</td>
<td>26.3</td>
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<tr>
<td>Berg et al. (1997)</td>
<td>345</td>
<td>48</td>
<td>28</td>
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<tr>
<td>Doxorubicin</td>
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<td>Berg et al. (1995)</td>
<td>256</td>
<td>50.5</td>
<td>9.7</td>
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<td>Moore et al. (2007)</td>
<td>240</td>
<td>35</td>
<td>17</td>
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<td>Cetuximab</td>
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<td>Bergman et al. (1996)</td>
<td>321</td>
<td>35.4</td>
<td>NR</td>
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<tr>
<td>Phillips et al. (2009)</td>
<td>307</td>
<td>36.8</td>
<td>18.7</td>
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<tr>
<td>Doxorubicin + Cetuximab</td>
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<tr>
<td>Bacon et al. (2008)</td>
<td>258</td>
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<td>Bailey et al. (2003)</td>
<td>235</td>
<td>NR</td>
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<tr>
<td>Keir et al. (2004a)</td>
<td>320</td>
<td>48</td>
<td>18</td>
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<tr>
<td>Cetuximab + Cetuximab + Protacuum</td>
<td>450</td>
<td>NR</td>
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<td>Langova et al. (2004)</td>
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<td>Ilapatinib</td>
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<td>Kipperstein et al. (2002b)</td>
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<td>31</td>
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NR, not reported.
However, when combined with chemotherapy, pamidronate did not improve pain alleviation (Fan et al., 2009). Some success in providing pain palliation in dogs with metastatic appendicular OSA has been achieved using metronomic chemotherapy with doxycycline, piroxicam and cyclophosphamide (Liptak et al., 2004b).

**Humans**

In humans, multimodality treatment is recommended for OSA. For high grade OSAs, preoperative (neoadjuvant) chemotherapy, wide surgical resection and post-operative chemotherapy are used. The most effective chemotherapeutic agents are doxorubicin, cisplatin, methotrexate and ifosfamide. Combined treatment serves to avoid chemoresistance and to increase the degree of tumour necrosis. There is usually a delay of 3-4 weeks after the last administration of neoadjuvant chemotherapy before the tumour is excised. Adjuvant chemotherapy is usually started 2 weeks after surgery.

Tumour necrosis after administration of preoperative chemotherapy is an important factor in determining the post-operative chemotherapy regimen (O’Day and Gorlick, 2009). Patients with P90% tumour necrosis at the time of surgery (good responders) will usually receive the same treatment regimen post-operatively as pre-operatively. In patients with <90% tumour necrosis (poor responders), post-surgery treatment usually includes a salvage regimen, either an increased dose or duration of the same chemotherapeutic agents or a different protocol, but neither have been shown to improve survival (Messerschmitt et al., 2009).

Amputation and limb-sparing procedures are the two principal surgical options in humans. No significant differences in survival rates and local recurrence (2.8-6%) are reported when amputation or limb-sparing surgery are used. Limb salvage is possible in more than 85% of human appendicular OSAs (Federman et al., 2009; Messerschmitt et al., 2009), but is contraindicated when resection with wide surgical margins is not feasible, in cases of neurovascular involvement or with pathological fractures. The options available for limb salvage include tumour removal and endoprosthetic replacement, rotationplasty, allografts and autografts. Limb-sparing complications include infection (11%) and implant failure. Tumour excision usually includes resection of both primary and metastatic sites; excision of all clinically detectable tumours is associated with a 5-fold increase in survival compared with excision of the primary tumour alone.

In humans, 5-year survival of 10-20% has been reported after amputation without chemotherapy, whereas a 5-year survival rate of 60-78% has been reported for non-metastatic patients when surgery is combined with systemic multi-agent chemotherapy (Malawer et al., 2008). Despite multimodality treatment, 30-40% of OSA patients still experience relapses within 3 years of treatment.

An increase in 5-year survival has been obtained by combining L-MTP-PE immunotherapy with chemotherapy in non-metastatic patients. Radiation therapy is mainly used as palliation for unresectable tumours, as well as for incompletely resected tumour excision margins (Federman et al., 2009). Pain palliation has also been achieved by use of radiopharmaceuticals such as 125I-Samarium (O’Day and Gorlick, 2009). Extracorporeal irradiation, including stereotactic radiosurgery or surgically exposed, irradiated and reimplanted bone, are among the more innovative and promising curative-intent uses of radiation therapy (Federman et al., 2009; Ta et al., 2009).

**Prognostic factors**

**Dogs**

Negative prognostic indicators associated with a shorter survival time in dogs with appendicular OSA include young age (Spodnick et al., 1992; Loukopoulos and Robinson, 2007), elevated pre-treatment total and bone-specific serum ALP activities (Ehrhart et al., 1998; Garzotto et al., 2000; Kirpenstein et al., 2002a), metastatic spread to regional lymph nodes (Hillers et al., 2005), high histological grade (grade III) (Kirpenstein et al., 2002a; Loukopoulos and Robinson, 2007), stage III OSA (distant metastases to bone and/or other sites) (Baccam et al., 2006), proximal humeral, rib or scapular involvement (Bergman et al., 1996), higher body weight (Hammer et al., 1995; Ru et al., 1998), incompleteness of excision (Hammer et al., 1995) and tumour volume (Misdorp and Hart, 1979). Percent tumour necrosis induced by chemotherapy or radiation therapy is predictive of local tumour control (Powers et al., 1991).

**Humans**

In humans, negative prognostic factors include metastases at presentation, metastatic spread to lymph nodes, poor response to preoperative chemotherapy, large tumour volume, increased activities of ALP and lactate dehydrogenase (LDH) in serum, primary localisation in the axial skeleton and inadequate surgical margins (Bacci et al., 2006; Malawer et al., 2008; Messerschmitt et al., 2009; Ta et al., 2009). In one study, time to relapse was longer in patients treated by neoadjuvant chemotherapy than in those given adjuvant chemotherapy (Ta et al., 2009).

**Conclusions**

Dogs with OSA represent a unique model for OSA in humans due to their similar clinical presentation and molecular features, along with the relatively high number of dogs diagnosed with OSA each year. Differences and similarities between human and canine OSA are summarised in Table 2. Improvements in diagnostic and imaging techniques, chemotherapy and surgical procedures have improved outcomes in both human and canine patients. However, there is still a need for effective
treatment of OSA, mainly to control metastatic disease. Important comparative advances have been made in the study of tumour biology and progression, risk factors and the evaluation of novel cancer strategies. There is likely to be increasing interest from the human cancer drug industry in conducting clinical trials in dogs with OSA before or concomitantly with trials in human patients.
## Table 2. Similarities and differences between human and canine appendicular osteosarcomas.

<table>
<thead>
<tr>
<th>Variables</th>
<th>Dog</th>
<th>Human</th>
</tr>
</thead>
<tbody>
<tr>
<td>Incidence in USA</td>
<td>Middle-aged to older dogs</td>
<td>Adolescent disease</td>
</tr>
<tr>
<td>Median age</td>
<td>Peak of incidence 7-9 years</td>
<td>Peak of incidence at 10-20 years</td>
</tr>
<tr>
<td>Race/breed</td>
<td>Second small peak at 18-24 months</td>
<td>Median peak age at 16 years</td>
</tr>
<tr>
<td>Sex</td>
<td>Median peak age at 17 years</td>
<td>None</td>
</tr>
<tr>
<td>Site</td>
<td>Males more than females: Ratio 1.5:1</td>
<td>Males or diaphysis of long bones (80-90%)</td>
</tr>
<tr>
<td>Aetiology</td>
<td>75% appendicular skeleton, metaphysis of long bones, mainly distal radius, proximal humerus, distal femur and proximal and distal tibia</td>
<td>Proximal humerus (25%)</td>
</tr>
<tr>
<td>Histopathological grade</td>
<td>Not completely known</td>
<td>Not completely known</td>
</tr>
<tr>
<td>Molecular and cellular genetic alterations</td>
<td>High grade</td>
<td>High grade</td>
</tr>
<tr>
<td>IGF-1/IGF-1R, IGF-1 receptor</td>
<td>p53: Mutated</td>
<td>p53: Mutated</td>
</tr>
<tr>
<td>IGFR1, IGF-1 receptor</td>
<td>Over-expressed; Poor clinical outcome</td>
<td>Over-expressed; Poor clinical outcome</td>
</tr>
<tr>
<td>HGF, hepatocyte growth factor, HGF/CGF Met: Over-expressed; Contributing to malignant phenotype</td>
<td>PDGFR-α, PDGFR-β, PDGF-α receptor</td>
<td>PDGF-B, PDGF-B receptor</td>
</tr>
<tr>
<td>ERBB-2/HER-2: Over-expressed; Poor clinical outcome</td>
<td>Mutated or down-regulated</td>
<td>Mutated or down-regulated</td>
</tr>
<tr>
<td>ERBB-3/HER-3: Over-expressed; Poor clinical outcome</td>
<td>Contributing to malignant phenotype</td>
<td>Contributing to malignant phenotype</td>
</tr>
<tr>
<td>Matrix metalloproteinases: Expressed</td>
<td>Matrix metalloproteinases: Expressed</td>
<td>Matrix metalloproteinases: Expressed</td>
</tr>
<tr>
<td>PDGFR-α, PDGFR-β, PDGF-B, PDGF-B receptor</td>
<td>Expressed</td>
<td>Expressed</td>
</tr>
<tr>
<td>VEGF, VEGF receptor</td>
<td>Expressed</td>
<td>Expressed</td>
</tr>
<tr>
<td>Clinical signs</td>
<td>Pain</td>
<td>Pain</td>
</tr>
<tr>
<td>Swelling</td>
<td>Leg swelling</td>
<td>Leg swelling</td>
</tr>
<tr>
<td>Hard painful mass</td>
<td>Hard painful mass</td>
<td>Hard painful mass</td>
</tr>
<tr>
<td>Uncommon pathological fracture (3%)</td>
<td>Uncommon pathological fracture</td>
<td>Uncommon pathological fracture</td>
</tr>
<tr>
<td>Metastatic site</td>
<td>10% of cases with metastasis at diagnosis: Lung, bone (7-8%)</td>
<td>20% of cases with metastasis at diagnosis: Lung, bone</td>
</tr>
<tr>
<td>Regional lymph node metastasis (4.4-9.0%)</td>
<td>Regional lymph node metastasis &lt; 10%</td>
<td>Regional lymph node metastasis &lt; 10%</td>
</tr>
<tr>
<td>Treatment</td>
<td>Amputation</td>
<td>Amputation (rare)</td>
</tr>
<tr>
<td>Limb-sparing techniques</td>
<td>Neoadjuvant chemotherapy: Doxorubicin, methotrexate, ifosfamide, platinum and adjuvant post-surgery</td>
<td></td>
</tr>
<tr>
<td>Adjuvant chemotherapy: Doxorubicin, platinum</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Survival</td>
<td>50% survival at 1 year with chemotherapy</td>
<td>70% survival at 5 years with chemotherapy</td>
</tr>
<tr>
<td>Negative prognostic indicators</td>
<td>Metastasis at diagnosis: Lung, bone, lymph nodes</td>
<td>Metastasis at diagnosis: Lung, bone, lymph nodes</td>
</tr>
<tr>
<td>High serum ALP, LDH activities</td>
<td>High serum ALP, LDH activities</td>
<td>High serum ALP, LDH activities</td>
</tr>
<tr>
<td>Tumour volume</td>
<td>Tumour volume</td>
<td>Tumour volume</td>
</tr>
<tr>
<td>Age: Young dogs</td>
<td>Age: Youngest affected</td>
<td>Age: Youngest affected</td>
</tr>
<tr>
<td>Positive prognostic indicators</td>
<td>Post-operative limb-sparing infection</td>
<td>Post-operative limb-sparing infection</td>
</tr>
<tr>
<td>High percentage of tumor necrosis induced by chemotherapy or radiotherapy</td>
<td>High percentage of tumor necrosis induced by chemotherapy or radiotherapy</td>
<td>High percentage of tumor necrosis induced by chemotherapy or radiotherapy</td>
</tr>
</tbody>
</table>

IGF-1, insulin-like growth factor-1; IGF-1R, IGF-1 receptor; HGF, hepatocyte growth factor, HER-2, human epidermal growth factor receptor 2; PTEN, phosphatase and tensin homolog, PDGF-B, platelet-derived growth factor-B; VEGF, vascular endothelial growth factor; P-gp, P-glycoprotein; ALP, alkaline phosphatase; LDH, lactate dehydrogenase.

### Conflict of interest statement

None of the authors of this paper has a financial or personal relationship with other people or organisations that could inappropriately influence or bias the content of the paper.
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